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(54) Title: COMPOSITIONS FOR REGENERATING TISSUE THAT HAS DEGENERATED, AND METHODS FOR USING SUCH COMPOSITIONS (57) Abstract A composition for promoting regeneration of tissue which has degenerated in a subject as a result of a disease or disorder and a method of using the composition is provided. The composition comprises a biodegradable acellular matrix, and passaged autologous fibroblasts substantially free of immunogenic proteins integrated within the matrix. The method of using the composition to promote regeneration of tissue involves placing the composition on a site of degenerated tissue in a subject so that the composition promotes tissue regeneration at the site. The composition and the method of its use have applications promoting regeneration of tissue that has degenerated as a result of numerous diseases or disorders, including, but not limited to, defects of the oral mucosa, trauma to the oral mucosa (e.g., extraction of a tooth), periodontal disease, diabetes, cutaneous ulcers, venous stasis, scars of the skin, or wrinkles of the skin. Also provided is an injectable composition comprising any type of collagen and passaged autologous fibroblasts substantially free of immunogenic proteins, for correcting defects in skin, such as wrinkles or scars, and for augmenting tissue in the subject, particularly facial tissue.		

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- 1 -

COMPOSITIONS FOR REGENERATING TISSUE THAT HAS
DEGENERATED, AND METHODS FOR USING SUCH COMPOSITIONS

5 Field of the Invention

The present invention concerns the regeneration of tissues in a subject that have degenerated as a result of a disease or disorder in the subject. More particularly, the present invention concerns novel compositions for use
10 in nonsurgical techniques that promote regeneration of tissue whose mass has been diminished due to a disease or disorder in a subject, correct defects in the skin of subjects, or augment tissue in subjects. Also disclosed is the use of a novel composition in conjunction with a
15 biodegradable acellular matrix for ameliorating defects in the tissues, and methods for using the novel composition.

Background of the Invention

"Periodontal disease" is the term commonly used to
20 describe inflammatory disease of the periodontium, i.e., the tissue surrounding and securing teeth to the jawbone. The condition is characterized by inflammatory and degenerative processes that develop at the gingival margin (gingivitis) and lead to a progressive breakdown
25 and resorption of the periodontal ligament and bone (periodontitis), oftentimes resulting in severe diminution of the periodontium. Periodontal disease is the leading cause of tooth loss in adults after middle age. [Anderson's Pathology, p. 2000, John M. Kissane ed.,
30 9th ed. (1992)].

- 2 -

Periodontal disease results from the accumulation of bacterial plaque in the gap between the gingiva and the tooth. While anaerobic bacteria are the primary etiologic agents, the destructive process is believed to be mediated in large part by immunologic reactions of the host. As the disease progresses, a periodontal pocket is established below the gingival margin, thus prolonging and promoting the inflammatory process. Successive inflammatory reactions result in the progressive erosion of the tooth-supporting tissues, i.e., the collagenous fibers making up the periodontal ligament and the bone pocket in which the tooth sits. [Reviewed in Anderson's Pathology, pp. 1999-2000, John M. Kissane ed., 9th ed. (1992); Shafer et al., A Textbook of Oral Pathology, 4th ed. (1983)].

Periodontal disease can be diagnosed by checking the gingiva for inflammation, probing the depths of periodontal pockets, checking clinical attachment level, and assessing bone loss by means of autoradiography. [Jeffcoat, M.K., et al., J. Am. Dent. Assoc., 128:713-724 (1997)].

A number of techniques, both surgical and nonsurgical, have been developed to treat periodontal disease. Particularly with respect to severe periodontitis, none of the currently available treatments are wholly satisfactory.

For relatively mild cases of periodontitis, practitioners have traditionally employed nonsurgical mechanical debridement (i.e. scaling and root planing) to remove the bacterial plaque whose accumulation perpetuates the disease, thereby reducing inflammation. Mechanical debridement can be accomplished using manual, sonic or ultrasonic instruments. Scaling and root planing have been shown to decrease gingival inflammation, decrease probe depth, and promote

- 3 -

maintenance of clinical attachment level. However, without resorting to surgical procedures, access to root surfaces and bony defects is restricted, and only limited debridement is possible. [Jeffcoat, M.K., et al., J. Am. Dent. Assoc. 128:713-724 (1997)].

As a result, nonsurgical scaling and root planing is insufficient to treat more severe cases of periodontitis, and it is necessary to resort to more aggressive surgical techniques. Surgical techniques comprise reflecting the gingival tissues to provide access to root surfaces and bone defects, in order that mechanical debridement may be accomplished directly. Following debridement, the gingival tissue is sutured back in position. Currently available surgical approaches entail substantial patient discomfort and fail to consistently provide satisfactory outcome.

There are a number of non-surgical, non-mechanical approaches to treating periodontal disease, including supragingival and subgingival irrigation and the application of chemical and antimicrobial agents. Yet none of these approaches have achieved more than marginal success [Jeffcoat, M.K., et al., J. Am. Dent. Assoc., 128:713-724 (1997)]. In particular, there are a number of deleterious side effects associated with the use of antibiotics, along with risks such as drug sensitivity and the emergence of antibiotic-resistant pathogens.

Another approach to combating destruction of the periodontal tissue has focused on inhibiting the matrix metalloproteases responsible for this destruction. Tetracyclines in particular have shown promise as inhibitors of extracellular collagenases, but cause the same side effects associated with antibiotics in general. Modified forms of tetracycline have been developed which are non-antimicrobial and retain their ability to inhibit collagenases, but these chemically modified tetracyclines

- 4 -

are not commercially available. [Ciancio, G.C. et al., J. Am. Dent. Assoc. 123:34-43 (1992)].

Because cell proliferation, cell migration and matrix synthesis are prerequisites for periodontal
5 regeneration, some researchers have attempted to use tissue growth factors, for example insulin-like growth factor, platelet-derived growth factor, and transforming growth factor to promote periodontal regeneration.

In summary, none of the nonmechanical approaches
10 to treating periodontitis have been able to offer more than modest, short term enhancement of traditional mechanical debridement. As noted in a 1997 review of techniques used in treating periodontal disease, "scaling and root planning accompanied by oral hygiene procedures
15 remains the first mode of treatment for adult periodontitis." [Jeffcoat, M.K. et al., J. Am. Dent. Assoc., 128:713-724 (1997)].

A great deal of research has been directed to methods of regenerating periodontal tissue lost as a
20 consequence of periodontal disease, but as yet no wholly satisfactory method is available. For the most part, efforts have focused on surgical approaches that fill the defects with a variety of materials (bone grafting) or use guided tissue regeneration.

25 Bone grafting techniques involve the use of natural bone or synthetic bone materials. Natural bone grafts are typically either autografts (grafts transferred from one position in the body of a patient to another position in the body of the same patient) or
30 allografts (grafts transferred from one person to another). Clinicians using natural bone grafts have had limited success in inducing new bone growth. Problems associated with the use of autografts include the need for a second surgical site and, in some cases, fresh

- 5 -

grafts may be associated with root resorption. [Jeffcoat, M.K. et al., J. Am. Dent. Assoc. 128:713-724 (1997)].

Freeze-dried, demineralized bone has been used as an allograft and shown to promote bone formation.

5 However, the predictability and the amount of bone fill achieved varies. [Jeffcoat, M.K. et al., J. Am. Dent. Assoc., 128:713-724 (1997)]. Since allografts are transferred from one person to another, the potential exists that viruses or other pathogens might be
10 transferred to the patient.

Synthetic bone materials which have been investigated include plaster, calcium carbonates, and ceramics such as hydroxyapatite. Clinical trials have demonstrated that the use of synthetic grafts has
15 resulted in improvements in probing depth and attachment level. Histologic findings, however, indicate that, in general, synthetic grafts act primarily as space fillers, with little if any regeneration. [Jeffcoat, M.K. et al., J. Am. Dent. Assoc., 128:713-724 (1997)].

20 Guided tissue regeneration is a surgical approach based on placing a membrane barrier under a soft tissue flap above the area of bone loss to enhance wound healing potential. [Ciancio, G.C. et al., J. Am. Dent. Assoc., 123:34-43 (1992)]. Investigators have studied both
25 resorbable and nonresorbable membranes. A significant disadvantage of using a nonresorbable membrane is the requirement of a second surgical procedure after approximately six weeks to remove the membrane. While less evidence is available for resorbable membranes than
30 for nonresorbable membranes, improvements in clinical attachment levels have been shown for both types of membranes compared with debridement alone. Most favorable results are reported for Class II furcations in the mandible and for intrabony defects. Less favorable
35 results have been reported in maxillary molar and Class

- 6 -

III (through and through) furcation defects. [Jeffcoat, M.K. et al., J. Am. Dent. Assoc., 128:731-724 (1997)].

In summary, none of the currently available treatments for periodontal disease is wholly
5 satisfactory, particularly with regards to regenerating periodontal tissue lost as a result of periodontitis.

The oral mucosa is the tissue lining the oral cavity. There are a number of conditions that can result in defects in the oral mucosa, for example trauma,
10 dermatoses, recurrent aphthous stomatitis, and infections. [Flint, S., The Practitioner 235:56-63 (1991)]. There is currently no wholly satisfactory means of correcting these defects.

Furthermore, there have been efforts to develop
15 and use compositions and methods to correct defects in skin, such as scars and wrinkles, or to augment the tissue of a subject in order to improve the appearance of the skin, particularly facial skin. The principal method employed to correct such defects involves injecting a
20 filler composition into the dermal layer of the skin proximate to the defect or desired tissue augmentation. Examples of non-biological filler compositions used in these roles include mineral oil, paraffin, silicone fluid, autologous fat, gelatin powder mixes,
25 polymethylmethacrylate microspheres, cross-linked polydimethylsiloxane, "TEFLON" paste, reconstituted bovine collagen, and autologous human collagen.

However, the use of these compositions comprises inherent limitations. For example, the use of mineral
30 oil, paraffin and similar oils and waxes has resulted in complications such as local chronic edema, lymphadenopathy, scarring and ulcerations (Devore et al., *Effectiveness of injectable filler materials for smoothing wrinkle lines and depressed scars. Medical*

- 7 -

Progress Through Technology 20:243-250 (1994 which is hereby incorporated by reference in its entirety).

The use of reconstituted bovine collagen to correct defects or augment tissue also possesses inherent limitations. For example, it has been reported that reconstituted bovine collagen is only moderately effective, and is associated with infrequent, but controversial, adverse reactions. In addition, it is rapidly broken down and resorbed in vivo, providing only a temporary correction of a skin defect or augmentation. More importantly, reconstituted bovine collagen may elicit an immune response in the subject. Id.

As explained above, gelatin matrix implant such as that sold under the mark "FRIBEL", is a composite material of porcine gelatin powder and o-aminocaproic acid which are dispersed in 0.9% (by volume) sodium chloride solution and an aliquot of the recipient's plasma mixed in a 1:1 ratio, is also used to correct skin defects and augment tissue. However, this material also possess inherent limitations. Specifically gelatin matrix does not appear to have applications in the treatment of wrinkle lines. Moreover, since a large bore needle (27 gauge or greater) is used to inject the gelatin matrix into the subject's skin, treatment with gelatin results in greater discomfort and pain to the subject as opposed to the injection of other fillers. Id.

In addition, the use of autologous fat injections to correct a skin defect or augment tissue in a subject, while eliminating the potential of eliciting an immune response, also possesses disadvantages. More specifically, prior to its injection, fat must be processed by skilled clinicians in aseptic conditions to maintain sterility. Also, a very large bore needle (as large as 22 gauge) is needed to inject the fat into a subject, resulting in great pain, moderate bruising, and

- 8 -

formation of visible puncture holes. Moreover, fat injections are subject to rapid resorption, and must be repeated in order to maintain skin augmentation or defect correction.

5 The use of autologous, injectable dermal collagen to correct defects or augment tissue has also met with limited success. For example, if large concentrations of collagen are injected, a 27 gauge needle or larger is used, resulting in the infliction of pain on the subject.
10 Furthermore, serial injections are required in order to compensate for the gradual resorption of autologous collagen.

Hence, what is needed is an efficient non-surgical composition that promotes the regeneration of tissues of
15 the gums or the palate and bone that have degenerated as a result of periodontal disease or trauma.

Moreover, what is needed is a composition and method for promoting regeneration of tissue that does not elicit an immune response in the subject at the site of
20 desired tissue regeneration.

What is also needed is a composition that can be used in non-surgical methods to correct defects in skin, such as scars or wrinkles, and augment tissue in a subject, particularly facial tissue, which is not rapidly
25 resorbed by the body so that additional injections are required.

U.S. patent no. 5,591,444, U.S. patent no. 5,660,850, and U.S. patent no. 5,665,372 are incorporated herein by reference in their entirety.

30 Summary of the Invention

The present invention provides a composition for promoting regeneration of tissue in a subject that has degenerated as a result of a disease or disorder, and a method of using the composition that does not suffer from

- 9 -

the shortcomings of other methods described above. The present invention is based on the inventors' discovery of the successful use of autologous fibroblasts to regenerate tissue in a subject, correct skin defects in the subject, or augment tissue in the subject.

5 Fibroblasts are connective-tissue cells involved in tissue repair. When a tissue is injured, nearby fibroblasts migrate into the wound, proliferate, and produce large amounts of collagenous matrix, which helps
10 to isolate and repair the damaged tissue. [Alberts et al., *Molecular Biology of the Cell*, p. 987, 2nd ed., (1992)].

Broadly, the present invention extends to a method for regenerating a subject's tissue that has degenerated
15 as a result of a disease or disorder, comprising the steps of providing a pharmaceutical composition comprising autologous, passaged fibroblasts, identifying a site of tissue degeneration, and injecting an effective amount of the composition into tissue at the site of
20 tissue degeneration so that the tissue is augmented and regeneration of tissue is promoted.

Injection of a pharmaceutical composition of the present invention can be into tissues of the subject comprising the periodontal pocket and/or the periodontal
25 tissue adjacent to the area of degeneration or into tissue subadjacent to a defect in the oral mucosa, or into the tissue of the palate of a human subject, in order promote regeneration of tissue in the oral mucosa, the gingiva, or the palate. Typical defects in the oral
30 mucosa or palate that can be corrected with this embodiment of the present invention include those caused by trauma, dermatoses, recurrent aphthous stomatitis, and infections, or a disease or disorder. Moreover, the present invention can be used to correct defects in the
35 skin, such as scars or wrinkles.

- 10 -

A disease or disorder which results in tissue degeneration in a subject that can be treated with the present invention includes, but is not limited to, defects of the oral mucosa, periodontal disease; trauma
5 to the oral mucosa (e.g., extraction of a tooth), diabetes, cutaneous ulcers, or venous stasis. Moreover, periodontal disease can include periodontal degeneration, gingivitis, or a non-healing wound of the palatal mucosa or the gingival mucosa.

10 The present invention further extends to a method of forming a composition comprising autologous, passaged fibroblasts which are substantially free of immunogenic proteins, and are histocompatible with a subject. This method comprises the steps of collecting a biopsy of
15 dermis from a subject, isolating the autologous fibroblasts contained in the biopsy from extracellular matrix and other cells contained in the biopsy, culturing the autologous fibroblasts in a culture medium that permits expansion of the autologous fibroblasts,
20 incubating the autologous fibroblasts in a protein free medium for at least about 2 hours between about 30°C and about 37.5°C, and exposing the incubated autologous fibroblasts to a proteolytic enzyme so as to suspend the fibroblasts. An example of a culture medium that permits
25 expansion of autologous fibroblasts comprises between 0.0% and about 20% serum, wherein the serum can be either human or non-human. Also, the biopsy from dermis can comprise tissue from the gums, palate or skin of the subject. Hence autologous fibroblasts from the gums,
30 plate or skin have applications in the present invention.

In another embodiment, the passaged autologous fibroblasts can be added to a pharmaceutically acceptable carrier to form a pharmaceutical composition. In
producing a pharmaceutical composition of the present
35 invention, antigenic proteins are removed from the

- 11 -

autologous fibroblasts, thereby avoiding an immunological reaction in a subject when such cells are reintroduced to the subject proximate to the site of tissue degeneration or defect.

5 In addition, the present invention extends to a device delivering a pharmaceutical composition of the present invention to a site proximate to the site of tissue degeneration or defect in a subject, wherein the device comprises a hypodermic syringe having a syringe
10 chamber, a piston disposed therein, an orifice communicating with the chamber, a pharmaceutical composition comprising autologous passaged fibroblasts and a pharmaceutically acceptable carrier, such that the pharmaceutical composition is disposed in the chamber,
15 and a hypodermic needle is fixed to the orifice.

Tissues which have suffered degeneration or have a defect that can be treated with a device of the present invention include the oral mucosa, the gingival mucosa, and the palatal mucosa. Moreover, diseases or disorders
20 which can be treated with this device include periodontal degeneration, gingivitis, or a non-healing wound of the palatal mucosa or the gingival mucosa. Also skin defects, such as scars and wrinkles, can be treated with the present invention.

25 The present invention further extends to a composition for promoting the regeneration of tissue that has degenerated in a subject, wherein the composition comprises a biodegradable acellular matrix, and autologous passaged fibroblasts substantially free of
30 immunogenic proteins, wherein the autologous fibroblasts are integrated into the biocompatible biodegradable acellular matrix. In an embodiment of the present invention, the biocompatible biodegradable acellular matrix comprises exogenous proteins, such as any type of
35 collagen. In addition, the biodegradable acellular

- 12 -

matrix can be comprised of any type of collagen and glycosaminoglycans (GAG) cross-linked with, for example, glutaraldehyde, or any type of collagen.

In yet another example, the biodegradable
5 acellular matrix comprises one or more of gelatin, polyglycolic acid, cat gut, demineralized bone, or hydroxyapatite. Other appropriate matrices consist of bone from which substantially all (e.g., at least 80%, at least 90%, at least 95%, at least 99%, or even 100% by
10 weight) organic material has been removed (referred to herein as "anorganic bone"); such matrices can, optionally, include exogenous collagen in various amounts (e.g., about 1%, about 2%, about 5%, about 10%, or about 20% by dry weight).

15 Also, diseases, disorders or defects resulting in degeneration of tissue in a subject which can be treated with the present invention, comprise defects of the oral mucosa, trauma to the oral mucosa (e.g., extraction of a tooth), periodontal disease, diabetes, cutaneous ulcers,
20 or venous stasis. In addition, examples of periodontal disease which result in tissue degeneration include, but are not limited to, periodontal degeneration, gingivitis, or non-healing wounds of the palatal mucosa or gingival mucosa. Moreover, defects in skin, such as scars or
25 wrinkles, can be treated with the present invention.

The present invention further extends to a method of using a composition for promoting regeneration of tissue, wherein the method comprises passaged autologous fibroblasts substantially free of immunogenic proteins,
30 providing a biodegradable acellular matrix, incubating the biodegradable acellular matrix with the passaged autologous fibroblasts so that the fibroblasts integrate into the biodegradable acellular matrix, forming a composition for promoting regeneration of tissue,
35 identifying a site of tissue degeneration or defect in

- 13 -

the subject, and applying the composition to the site of tissue degeneration or defect so that tissue regeneration at the site is promoted. Autologous passaged fibroblasts used herein can comprise fibroblasts from the gums,
5 palate or skin of the subject.

Diseases or disorders which can be treated with this method include, but are not limited to, defects of the oral mucosa, trauma to the oral mucosa (e.g., extraction of a tooth), periodontal disease, diabetes,
10 cutaneous ulcers, or venous stasis. Examples of periodontal disease that can be treated with the present invention comprise periodontal degeneration, gingivitis, or a non-healing wound of the palatal mucosa or the gingival mucosa. Moreover, defects in skin such as scars
15 or wrinkles can be treated with the composition of the present invention. In a preferred embodiment, such defects are treated with a composition comprising fibroblasts from the palate.

Furthermore, biodegradable acellular matrices
20 having applications in the present invention may comprise exogenous proteins. Examples of such matrices include matrices comprising any type of collagen, or any type of collagen and glycosaminoglycans (GAG) cross-linked with, for example, glutaraldehyde.

25 Other examples of biodegradable acellular matrices having applications in the present invention include one or more of gelatin, polyglycolic acid, cat gut, demineralized bone, or hydroxyapatite. Other appropriate matrices consist of bone from which substantially all
30 (e.g., at least 80%, at least 90%, at least 95%, at least 99%, or even 100% by weight) organic material has been removed (referred to herein as "anorganic bone"); such matrices can, optionally, include exogenous collagen, in various amounts (e.g., about 1%, about 2%, about 5%,
35 about 10%, or about 20% by dry weight).

- 14 -

The present invention further extends to an injectable composition for correcting a defect in skin of a subject, or augmenting tissue of a subject, said injectable composition comprising passaged, autologous fibroblasts substantially free of immunogenic proteins, and a biodegradable, acellular injectable filler material. Passaged autologous fibroblasts having applications in an injectable composition of the present invention are from gums, palate or skin of the subject.

Furthermore, the present invention extends to an injectable composition as described above, wherein the biodegradable, acellular injectable filler material comprises endogenous proteins. In particular, the acellular injectable filler material of an injectable composition of the present invention comprises an injectable dispersion of autologous collagen fibers having a concentration in the composition of at least 24 mg/ml of composition.

In addition, the present invention extends to an injectable composition as described above, wherein the biodegradable acellular injectable filler material comprises exogenous proteins, such as any type of collagen. An example of an exogenous collagen having applications in an injectable composition of the present invention is reconstituted bovine collagen fibers cross-linked with glutaraldehyde.

Furthermore, the filler material can comprise any type of solubilized gelatin either alone, or in combination with other materials. In a particular example, the filler material comprises porcine gelatin powder and o-aminocaproic acid dispersed in sodium chloride solution and an aliquot of plasma from the subject to be injected with the composition. Preferably the ratio of sodium chloride to serum is 1:1 by volume. Other examples of materials having applications in the

- 15 -

present invention as biodegradable, acellular injectable filler material include, but are not limited to polyglycolic acid or cat gut.

The present invention further extends to a method
5 for correcting a defect in skin of a subject, or augmenting tissue of a subject, wherein the method comprises injecting an effective amount of an injectable composition comprising autologous passaged fibroblasts substantially free of immunogenic proteins and a
10 biodegradable, acellular injectable filler material, into the skin of the subject at the site of the skin defect or desired tissue augmentation, so that regeneration of tissue at the site is promoted at the site.

Moreover, the present invention extends to a
15 method for correcting a defect in skin of a subject, or augmenting tissue of a subject, the method comprising the steps of injecting autologous fibroblasts substantially free of immunogenic proteins into the subject at a site of a skin defect or desired tissue augmentation, and
20 subsequently injecting a biodegradable, acellular injectable filler material into the site. In a particular embodiment of this method of the present invention, the duration between injecting the autologous fibroblasts into the subject and injecting the
25 biodegradable acellular injectable filler into the subject is about two weeks.

Autologous fibroblasts having applications in methods of the present invention for correcting a defect in skin of a subject, or augmenting tissue of a subject
30 can be obtained from the gums, palate of skin of the subject.

The present invention further extends to a method for correcting a defect in skin of a subject, or augmenting tissue of the subject, as described above,
35 wherein the biodegradable, acellular injectable filler

- 16 -

material comprises endogenous proteins. For example, the biodegradable acellular injectable filler material can comprise an injectable dispersion of autologous collagen fibers, preferably at a concentration of at least 24 mg
5 of autologous fibers per ml of composition.

Furthermore, the present invention extends to a method for correcting a defect in skin of a subject, or augmenting tissue of the subject, as described above, wherein the biodegradable, acellular filler material of
10 the composition comprises exogenous proteins such as, for example, any type of collagen. An example of collagen having applications in a method of the present invention comprises reconstituted bovine collagen fibers cross-linked with glutaraldehyde.

15 Other examples of biodegradable, acellular injectable filler material for use in a method for correcting a defect in skin or a subject, or augmenting tissue of the subject include, but are not limited to solubilized gelatin, polyglycolic acid, or cat gut
20 sutures. More specifically, an example of acellular injectable filler material having applications in the present invention comprises porcine gelatin powder and aminocaproic acid dispersed in sodium chloride solution, and an aliquot of plasma from the subject. Preferably,
25 the ratio of sodium chloride solution to the aliquot of serum is 1:1 by volume. Furthermore, the sodium chloride solution comprises 0.9% sodium chloride by volume.

In addition, the present invention extends to a method for correcting a defect in skin of a subject, or
30 augmenting tissue of a subject, as described above, wherein the ratio of autologous fibroblasts substantially free of immunogenic proteins to biodegradable, acellular injectable filler material is approximately 1:1 by volume.

- 17 -

Accordingly, it is an object of the present invention to provide a composition for augmenting tissue, or promoting regeneration of tissue such as the oral mucosa, the gingival mucosa, or the palatal mucosa or
5 skin, which has degenerated as a result of a disease or disorder. Examples of such disorders include periodontal disease, trauma, dermatoses, recurrent aphthous stomatitis, infections, scars, or wrinkles.

It is another object of the present invention to
10 provide a composition for augmenting tissue, or promoting regeneration of tissue, wherein the composition is histocompatible with a subject, thereby avoiding elicitation of an immune response and inflammation in the tissues of the subject near the site of degeneration of
15 tissue.

It is yet another object of the present invention to provide method of promoting tissue regeneration that does not require surgery.

It is yet still another object of the present
20 invention to promote regeneration of tissue in a subject without the use of antibiotics in the subject, and hence prevent the emergence of antibiotic resistant pathogens and deleterious side effects associated with antibiotics in the subject.

It is another object to provide an injectable
25 composition for correcting defects in skin, such as scars or wrinkles, or for augmenting tissue in a subject, particularly facial tissue (such as lips), which includes passaged autologous fibroblasts that can withstand
30 resorption so that subsequent injections are not needed, and to prevent the elicitation of an immune response in the subject.

It is yet another object of the present invention to provide methods for correcting defects in skin, such
35 as scars or wrinkles, or for augmenting tissue in a

- 18 -

subject, that employs the injectable composition set forth above, that inflicts limited pain on the subject, and does not elicit an immune response in the subject, and is not rapidly resorbed.

- 5 Other objects and advantages will become apparent to those skilled in the art from a review of the ensuing description.

Detailed Description of the Invention

10 The present invention is based, in part, on the recognition that living cells normally present in tissue that has degenerated, particularly fibroblasts, are the ideal material to augment the volume of tissue in order to promote regeneration of tissue. Hence the present invention ameliorates and reverses the degenerative
15 effects of a disease or disorder which results in tissue degeneration.

 Additionally, the present invention is based on the recognition that an ideal composition with which to augment the oral mucosa subadjacent to a defect, to treat
20 defects in the palate or gingiva, or to promote regeneration of tissue that has degenerated as a result of periodontal disease, would comprise living cells normally present in such tissues, particularly fibroblasts.

25 Moreover, the present invention is based on the recognition that an abundant supply of autologous cells of the desired type can be obtained by culturing a biopsy specimen taken from the skin, palate or gums of a subject several weeks prior to treating the tissue degenerating
30 disease, disorder, or defect. The invention is further based on the recognition that, after such a tissue culture expansion, the autologous cells will contain a significant quantity of antigenic proteins, but that the

- 19 -

antigenic proteins can be removed, prior to treatment of the subject.

The term "biodegradable" as used herein denotes a composition that is not biologically harmful and can be
5 chemically degraded or decomposed by natural effectors (e.g., weather, soil bacteria, plants, animals).

The term "autologous" as used herein refers to cells removed from a donor and administered to a recipient, wherein the donor and recipient are the same
10 individual.

The term "effective amount" as used herein refers to the injection of an amount of pharmaceutical composition of the present invention to promote tissue regeneration in of tissue that has degenerated in a
15 subject.

EXAMPLE I: Administration of a Suspension of Autologous Fibroblasts to Promote Tissue Regeneration and Correct Defects in Tissues

20 Method of Obtaining an Injectable Cell Pharmaceutical Composition

As disclosed above, one embodiment of the present invention comprises a method for regenerating tissue that has been damaged in a subject as a result of a disease or disorder in the subject, wherein the method comprises
25 providing a pharmaceutical composition comprising of autologous, passaged fibroblasts substantially free of immunogenic proteins, identifying a site of tissue degeneration or a defect in tissue, injecting an effective amount of the pharmaceutical composition into
30 the tissue at the site of the tissue degeneration or defect so that the tissue is augmented, and the growth of tissue is promoted at the site of degenerated tissue.

- 20 -

A disease or disorder which promotes tissue degeneration in a subject, and can be treated with this aspect of the present invention includes, but is not limited to, defects of the oral mucosa, trauma to the oral mucosa, periodontal disease, diabetes, cutaneous ulcers, or venous stasis. Furthermore, examples of periodontal disease which can be treated with this aspect of the present invention include periodontal degeneration, gingivitis, or a non-healing wound of the palatal mucosa or the gingival mucosa. In addition, defects in skin, such as scars or wrinkles can be treated with the present invention.

The invention can be practiced by injecting any undifferentiated mesenchymal cell that can be expanded in culture. In a preferred embodiment, dermal fibroblasts are injected because they can be readily obtained and expanded, and because they are a cell type normally present beneath the gingival mucosa or palatal mucosa. Fibroblasts taken from a biopsy of the gums, palate, or skin of the subject can be used in the present invention.

In providing a composition of the present invention, a dermal fibroblast culture is initiated from a 1 to 5 mm full thickness biopsy specimen of the gums, palate or skin of a subject suffering from tissue degeneration. Because of the phenomenon of allograft rejection, which is well known to transplantation surgeons and immunologists, it is essential that the cultured fibroblasts be histocompatible with the host. Histocompatibility can be ensured by obtaining a biopsy of the subject to be treated and culturing the fibroblasts from this specimen.

Before the initiation of the culture, the biopsy is washed repeatedly with antibiotic and antifungal agents. The specimen of dermis is then separated into small pieces. The pieces of the specimen are

- 21 -

individually placed onto the dry surface of a tissue culture flask and allowed to attach for between about 5 and about 10 minutes, before a small amount of medium is slowly added, taking care not to displace the attached tissue fragments. After about 48 hours of incubation, the flask is fed with additional medium. When a T-25 flask is used to start the culture, the initial amount of medium is about 1.5-2.0 ml. The establishment of a cell line from the biopsy specimen ordinarily takes between about 2 and 3 weeks, at which time the cells can be removed from the initial culture vessel for expansion.

During the early stages of the culture, it is desirable that the tissue fragments remain attached to the culture vessel bottom; fragments that detach should be reimplanted into new vessels. The fibroblasts can be stimulated to grow by a brief exposure to EDTA-trypsin, according to techniques well known to those skilled in the art. The exposure to trypsin is too brief to release the fibroblasts from their attachment to the culture vessel wall. Immediately after the cultures have become established and are approaching confluence, samples of the fibroblasts can be processed for frozen storage, such as in liquid nitrogen. Presently, numerous methods for successfully freezing cells for later use are known in the art, and are included in the present invention. The frozen storage of early rather than late passage fibroblasts is preferred because the number of passages in cell culture of normal human fibroblasts is limited.

The fibroblasts can be frozen in any freezing medium suitable for preserving fibroblasts. A medium consisting of about 70% (v/v) growth medium, about 20% (v/v) fetal bovine serum and about 10% (v/v) dimethylsulfoxide (DMSO) can be used with good effect. DMSO can also be substituted with, for example, glycerol. Thawed cells can be used to initiate secondary cultures

- 22 -

to obtain suspensions for use in the same subject without the inconvenience of obtaining a second specimen.

Any tissue culture technique that is suitable for the propagation of dermal fibroblasts from biopsy
5 specimens may be used to expand the cells to practice the invention. Techniques well known to those skilled in the art can be found in R.I. Freshney, Ed., ANIMAL CELL CULTURE: A PRACTICAL APPROACH (IRL Press, Oxford, England, 1986) and R.I. Freshney, Ed., CULTURE OF ANIMAL
10 CELLS: A MANUAL OF BASIC TECHNIQUES, Alan R. Liss & Co., New York, 1987), which are hereby incorporated by reference.

The medium can be any medium suited for the growth of primary fibroblast cultures. The medium can be
15 supplemented with human or non-human serum in an amount of between about 0.0% and about 20% (v/v) to promote growth of the fibroblasts. Higher concentrations of serum promote faster growth of the fibroblasts. An example of medium having application herein comprises
20 glucose DMEM supplemented with about 2 mM glutamine, about 10 mg/L sodium pyruvate, about 10% (v/v) fetal bovine serum and antibiotics ("complete medium"), wherein the concentration of glucose ranges from about 1,000 milligrams per liter of medium to about 4,500 milligrams
25 per liter of medium. Fibroblasts can also be expanded in a serum-free medium.

Autologous fibroblasts can be passaged into new flasks by trypsinization. For expansion, individual flasks are split 1:3. Triple bottom, T-150 flasks,
30 having a total culture area of 450 cm² are suitable for the practice of the invention. A triple bottom T-150 flask can be seeded with about 1X10⁶ to about 3X10⁶ cells and has a capacity to yield about 8X10⁶ to about 1.3X10⁷ cells. When the capacity of the flask is reached, which
35 typically requires about 5-7 days of culture, the growth

- 23 -

medium is replaced by serum-free medium; thereafter the cells are incubated, i.e., held at between about 30°C and about 37.5°C, for at least 2 hours. The incubation of the cells in serum free medium substantially removes from the cells proteins that are derived from the fetal bovine serum which, if present, can elicit an immune response in the subject. In a preferred embodiment, serum-free medium comprises glucose DMEM supplemented with about 2 mM glutamine, and about 110 mg/L sodium pyruvate, wherein the concentration of glucose can range from approximately 1,000 mg/L of medium to about 4,500 mg/L of medium. In a preferred embodiment, the concentration of glucose is approximately 4,500 mg/L of medium.

At the end of the incubation in serum free medium, the cells are removed from the tissue culture flask by trypsin-EDTA; washed extensively by centrifugation and resuspension; and suspended for injection in an equal volume of injectable isotonic solution with an appropriate physiological osmolarity, which is substantially pyrogen and foreign protein free. An example of such an isotonic solution is isotonic saline. Five triple bottom T-150 flasks, grown to capacity, yield about 3.5×10^7 to about 7×10^7 cells which are sufficient to make up about 1.2 to about 1.4 ml of suspension. A pharmaceutically acceptable carrier can then be added to the passaged autologous fibroblasts forming a pharmaceutical composition. The phrase "pharmaceutically acceptable" refers to molecular entities and compositions that are not deleterious to the cells, are physiologically tolerable, and do not typically produce an allergic or similar untoward reaction, such as gastric upset, dizziness and the like, when administered to a human. Such compositions include diluents of various buffer content (e.g., Tris-HCl, acetate, phosphate), pH and ionic strength.

- 24 -

Alternatively, the cells can be transported at 4°C so long as they are injected within 24-48 hours of the time that the pharmaceutical composition is made. The cells can be suspended in an appropriate physiological solution with appropriate osmolarity and tested for pyrogen and endotoxin levels, except for the absence of phenol red pH indicator, and the replacement of the fetal bovine serum by the subject's serum for such transportation (transport medium). In another embodiment the cells can be suspended in Krebs-Ringer solution comprising 5% dextrose or any other physiological solution. The cells can be aspirated and injected in the transport medium.

The volume of saline or transport medium in which the cells are suspended is related to such factors as the number of fibroblasts the practitioner desires to inject, the extent of the damage due to tissue degeneration or defect, or the size, the number of the defects that are to be treated, and the urgency of the subject's desire to obtain the results of treatment. Moreover, the practitioner can suspend the cells in a larger volume of medium and inject correspondingly fewer cells at each injection site.

Device for Treating Degenerated Tissue with
25 Autologous Passaged Fibroblast Pharmaceutical Composition
of the Present Invention

As explained above, the present invention extends to a device delivering a pharmaceutical composition of autologous passaged cells described above, to a point proximate to the site of tissue degeneration, or defects of the palate, oral mucosa or skin. Such a device comprises a hypodermic syringe having a syringe chamber, a piston disposed therein, an orifice communicating with the chamber, a pharmaceutical composition comprising

- 25 -

autologous, passaged fibroblasts, and a pharmaceutically acceptable carrier thereof, such that the pharmaceutical composition is disposed in the chamber, and a hypodermic needle is fixed to the orifice.

5 The Administration of a Pharmaceutical Composition

A pharmaceutical composition of the invention can be used to treat tissue degeneration in a subject as a result of a disorder or disease, such as periodontal disease, or defects of the oral mucosa or skin, such as
10 scars or wrinkles, by use of the following techniques.

Initially, the tissue to be injected is prepped with alcohol and stretched to give a taut surface. If the tissue degeneration is the result of periodontal disease, the tissue to be injected is periodontal tissue,
15 including periodontal pockets. If the tissue to be injected contains defects in the palate or gums of the subject, the tissue to be injected with a pharmaceutical composition of the present invention is subadjacent to the defect. Also, if the tissue to be injected is the
20 skin, in order to treat defects such as scars or wrinkles, it is also injected subadjacent to the defect.

After the tissue to be injected has been prepped, a syringe is filled with a pharmaceutical composition of the present invention and fitted with a 30 gauge needle.
25 The needle is inserted into the tissue as superficially as possible, and the orientation of the bevel is not critical to the success of this method of the present invention. The injection of the pharmaceutical composition is made by gentle pressure until a slight
30 blanch is seen in the injected tissue. Multiple serial injections are made.

- 26 -

Example II: A Composition for Promoting the
Regeneration of Tissue that Has
Degenerated in a Subject

Also disclosed in the present invention is a
5 composition for promoting the regeneration of tissue that
degenerated in a subject. Such degeneration can occur as
a result of periodontal disease, trauma, dermatoses,
recurrent aphthous stomatitis, or infections to name only
a few. Moreover, examples of periodontal disease which
10 can cause tissue degeneration include periodontal
degeneration, gingivitis, or non-healing wounds of the
palatal mucosa or gingival mucosa. Moreover, a
composition of the present invention can also be used to
correct defects in tissue of a subject, such as, for
15 example, defects in the palatal mucosa, gingival mucosa,
or defects in skin, such as scars or wrinkles. Such
compositions can be used, for example, for healing
extraction sockets after extraction of a tooth. A
composition of the present invention comprises a
20 biodegradable acellular matrix, and autologous passaged
fibroblasts substantially free of immunogenic proteins,
wherein the autologous fibroblasts are integrated into
the biodegradable acellular matrix.

Moreover, disclosed herein is a method of making a
25 composition of the present invention to promote
regeneration of tissue in a subject. Such a method
comprises providing a suspension of passaged autologous
fibroblasts substantially free of immunogenic proteins,
providing a biodegradable acellular matrix, incubating
30 the biodegradable acellular matrix with the suspension of
passaged autologous fibroblasts such that the autologous
fibroblasts integrate within the biodegradable acellular
matrix forming a composition for promoting regeneration
of tissue. Also disclosed herein is a method of using a
35 composition of the present invention, comprising

- 27 -

identifying a site of tissue degeneration, and applying the composition for promoting regeneration of tissue to the site of tissue degeneration.

In order to produce a composition of the present invention, autologous passaged fibroblasts substantially free of antigenic proteins must be made available to avoid the phenomenon of allograft rejection, which is well known to transplantation surgeons and immunologists. Hence, it is essential that the cultured fibroblasts be histocompatible with the host. Histocompatibility can be ensured by obtaining a biopsy of the gums, palate or skin of the subject to be treated, and culturing the fibroblasts from this specimen.

Before the initiation of the dermal fibroblast culture, a biopsy of 1-5 mm is taken from the gums, palate or skin of the subject, and washed repeatedly with antibiotic and antifungal agents. The specimen of dermis is then separated into small pieces. The pieces of the specimen are individually placed onto a dry surface of a tissue culture flask and allowed to attach for between 5 and 10 minutes before a small amount of medium is slowly added, taking care not to displace the attached tissue fragments. After 48 hours of incubation, the flask is fed with additional medium. When a T-25 flask is used to start the culture, the initial amount of medium is 1.5-2.0 ml. The establishment of a cell line from the biopsy specimen ordinarily takes between 2 and 3 weeks, at which time the cells can be removed from the initial culture vessel for expansion.

During the early stages of the culture it is desirable that the tissue fragments remain attached to the culture vessel bottom; fragments that detach should be reimplanted into new vessels. The autologous fibroblasts can be stimulated to grow by a brief exposure to trypsin-EDTA, according to techniques known to those

- 28 -

skilled in the art. The exposure to trypsin is too brief to release the fibroblasts from their attachment to the culture vessel wall.

Immediately after the cultures have become
5 established and are approaching confluence, samples of the autologous fibroblasts can be processed for frozen storage in, for example, liquid nitrogen. The frozen storage of early rather than late passage fibroblasts is preferred because the number of passages in cell culture
10 of normal human fibroblasts is limited.

The autologous fibroblasts can be frozen in any freezing medium suitable for preserving cells. A medium consisting of about 70% growth medium, about 20% (v/v) fetal bovine serum and about 10% (v/v) dimethylsulfoxide
15 (DMSO) can be used with good effect. DMSO can also be substituted with, for example, glycerol. Thawed cells can be used to initiate secondary cultures to obtain suspensions for use in the same subject without the inconvenience of obtaining a second specimen.

20 Any tissue culture technique that is suitable for the propagation of dermal fibroblasts from biopsy specimens may be used to expand the cells to practice the invention. Techniques for propagation known to those skilled in the art can be found in R.I. Freshney, Ed.,
25 ANIMAL CELL CULTURE: A PRACTICAL APPROACH (IRL Press, Oxford England, 1986) and R.I. Freshney, Ed., CULTURE OF ANIMAL CELLS: A MANUAL OF BASIC TECHNIQUES, Alan R. Liss & Co., New York, 1987), which are hereby incorporated by reference.

30 The medium can be any medium suited for the growth of primary autologous fibroblast cultures. In most instances, the medium is supplemented with human or non-human serum in the amount of between about 0.0% and about 20% (v/v) to promote growth of the autologous
35 fibroblasts. Higher concentrations of serum promote

- 29 -

faster growth of the fibroblasts. In a preferred embodiment the serum is fetal bovine serum, which is added to a final concentration of about 10% of medium. Moreover, the medium can be glucose DMEM supplemented with about 2mM glutamine, about 110 mg/L sodium pyruvate, about 10% (v/v) fetal bovine serum and antibiotics ("complete medium"), wherein the concentration of glucose ranges from approximately 1,000 mg/L of medium to approximately 4,500 mg/L of medium. Preferably, the concentration of glucose in the medium is approximately 4,500 mg/L of medium.

Autologous fibroblasts can be passaged into new flasks by trypsinization. For expansion, individual flasks are split 1:3. Triple bottom, T-150 flasks, having a total culture area of 450 cm² are suitable for the practice of the invention. A triple bottom T-150 flask can be seeded with about 1X10⁶ to about 3X10⁶ to about 3X10⁶ cells and has a capacity to yield about 8X10⁶ to about 1.3X10⁷ cells. When the capacity of the flask is reached, which typically requires 5-7 days of culture, the growth medium is replaced by serum-free medium; thereafter the cells are incubated, i.e., held at between about 30°C and about 37.5°C, for at least 2 hours. The incubation of the cells in serum free medium substantially removes from the cells proteins that are derived from the fetal bovine serum which, if present, would elicit an immune subject. In a preferred embodiment, the serum-free medium comprises glucose and DMEM supplemented with about 2 mM glutamine, about 110 mg/L sodium pyruvate, wherein the concentration of glucose ranges from approximately 1,000 mg/L of medium to approximately 4,500 mg/L of medium, and preferably is 4,500 mg/L of medium.

A biodegradable acellular matrix is then provided. Examples of such matrices which can be used in the

- 30 -

present invention include, but are not limited to, acellular matrices comprising exogenous proteins, or matrices comprising biodegradable polymers.

Numerous biodegradable acellular matrices
5 comprising exogenous proteins are presently available, and have ready applications in the present invention. An embodiment of such biodegradable acellular matrices are matrices comprising any type of collagen, or any type of collagen with glycosaminoglycans (GAG) cross-linked with,
10 for example, glutaraldehyde. Examples of collagen matrices having application in the present invention are absorbable collagen sponges made by the Calcitek Company of Carlsbad, CA. These collagen sponge dressings, sold under the names COLLATAPE," "COLLACOTE," and "COLLAPLUG"
15 are made from cross-linked collagen extracted from bovine deep flexor (Achilles) tendon, and glycosaminoglycans (GAGS). These products are soft, pliable, nonfriable, and non-pyrogenic. In addition, more than 90% of this product consists of open pores. An alternative
20 biodegradable acellular matrix can consist of collagen (e.g., bovine porcine collagen type I) formed into a thin membrane. Such a membrane is manufactured by the Calcitek Company and is marketed as BioMend™. Another such membranous matrix manufactured by ED. GEISTLICH
25 SÖHNE AG of Wolhusen, Switzerland, is made of porcine type I and type III collagen and is marketed as Bio-Gide®. Bio-Gide® has a bilayer structure with one surface that is porous allowing the ingrowth of cells and a second surface that is dense and will prevent the
30 ingrowth of fibrous tissue. Another biodegradable acellular matrix can be made from bone spongiosa formed into granules or blocks. This material consists of animal (e.g., human, non-human primate, bovine, sheep, pig, or goat) bone from which substantially all organic
35 material (e.g., proteins, lipids, nucleic acids,

- 31 -

carbohydrates, and small organic molecules such as vitamins and non-protein hormones) have been removed. This type of matrix is referred to herein as an anorganic matrix. One such matrix, which is marketed as either

5 Bio-Oss® spongiosa granules or Bio-Oss® blocks, is manufactured by ED. GEISTLICH SÖHNE AG. This company also manufactures a block-type matrix (Bio-Oss® collagen), also consisting of the anorganic bone, but containing in addition approximately 10% by weight of

10 collagen fibers. Other biodegradable acellular matrices having applications in the present invention can contain one or more of gelatin, polyglycolic acid, or cat gut sutures, demineralized bone, hydroxyapatite, or mixtures of these substances. Demineralized bone can be combined,

15 for example, with collagen to produce a matrix in the form of a sponge, block, or membrane. Synthetic polymers made from 1 or more monomers can also be used to make the biodegradable acellular matrices of the invention. The matrices can be made from one or more of such synthetic

20 polymers. The synthetic polymers can also be combined with any of the above-mentioned substances to form matrices. Different polymers forming a single matrix can be in separate compartments or layers. For example, Gore-Tex, Inc. manufactures a porous biodegradable

25 acellular matrix (GORE RESOLUT XT Regenerative Material) that is composed of a synthetic bioabsorbable glycolide and trimethylene carbonate copolymer fiber (into which cells can migrate) attached to an occlusive membrane that does not permit ingrowth of cells and composed of a

30 synthetic bioabsorbable glycolide and lactide copolymer.

After a biodegradable acellular matrix has been selected, a concentrated suspension of autologous passaged fibroblasts is evenly distributed on the surface of the matrix. Using a concentrated suspension is

- 32 -

necessary to avoid going beyond the capacity of the matrix to absorb the suspension. For example, a typical distribution of cell suspension using the GORE RESOLUT XT as the matrix comprises applying about 94 μ l to about 125 μ l of cell suspension having about 2.0×10^6 cells to about 4.0×10^6 cells, per square centimeter of matrix. Cells are allowed to attach to the matrix without further addition of media. In a preferred embodiment, incubation of the cells with the matrix occurs at about 37°C for about 1-2 hours. After at least sixty minutes of incubation, the cells readily attach and grow into the matrix, and are supplemented with additional growth medium. Cells are then cultured in the matrix for about 3 to 5 days. The matrix with the cells is then washed at least three times, for 10 minutes per wash, with, e.g., phosphate buffered solution (PBS) to substantially remove immunogenic proteins which may elicit an immune response in the subject. Fresh PBS is used for each washing. The matrix is then incubated for at least one hour in fresh PBS prior to use. After incubation, the matrix comprising autologous fibroblasts is placed on the area of tissue degeneration, or defect, such as, for example, the periodontal pockets, and secured so that it can not be moved from the site.

In the case of sponge matrix (e.g. Collacote®), approximately $1.5 - 2.0 \times 10^7$ fibroblasts in approximately 1.5ml of growth medium are seeded onto 2cm x 4cm thin (approximately 2.5mm to 3.0mm thick) sponges. The sponge is then incubated at 37°C for about 1-2 hours without further addition of medium to allow substantially all the fibroblasts to adhere to the matrix material. After cell adherence, additional growth medium is added to the composition of matrix and fibroblasts which is then incubated at 37°C for 3-4 days (with daily change of medium) to allow for fibroblast cell division and,

- 33 -

consequently, an increase in the number of fibroblasts in the matrix. The composition is then removed from growth medium containing FBS and washed repeatedly (at least 3 times) with FBS-free PBS. After each addition of PBS, 5 the matrix is incubated for 10-20 minutes prior to discarding of the PBS. After the final wash, the composition is either applied directly to the area of the subject requiring tissue regeneration or is transferred to a shipping vial containing a physiological solution 10 (e.g., Kreb's Ringer solution) and shipped (preferably overnight) to a practitioner (e.g., a dentist or physician).

In the case of a membranous matrix (e.g. Biomend™), approximately $3-8 \times 10^6$ fibroblasts in 15 approximately 100 μ l of growth medium are seeded onto the 15mm x20mm thin (approximately 0.5 to 1.0mm thick) membranes. The membrane is then incubated at 37°C for about 30-60 minutes without further addition of medium to allow substantially all the fibroblasts to adhere to the 20 matrix material. After cell adherence, additional growth medium is added to the composition of matrix and fibroblasts which is then incubated at 37°C for 2-3 days (with daily change of medium) to allow for fibroblast cell division and, consequently, an increase in the 25 number of fibroblasts bound to the matrix. Washing of the composition and either immediate use or shipping are as described above for the sponge matrices.

In the case of a block matrix such as the above described anorganic matrix (e.g., Bio-Oss® blocks), 30 approximately $5-10 \times 10^6$ fibroblasts in approximately 100 μ l to 150 μ l of growth medium are seeded into 1cm x 1cm x 2cm cubic blocks of matrix material. Cells are slowly seeded onto one face of the block face. Once the medium and cells have been absorbed into the block, another face 35 of the block is seeded in a similar fashion. The

- 34 -

procedure is repeated until all faces of the block have been seeded and the block is fully saturated with medium. Care is taken to avoid adding excess medium and thereby causing leaking out of medium and cells from the block.

- 5 The composition is then incubated at 37°C for about 30-60 minutes without further addition of medium to allow substantially all the fibroblasts to adhere to the matrix material. After cell adherence, additional growth medium is added to the composition of matrix and
- 10 fibroblasts which is then incubated at 37°C for 2-3 days (with daily change of medium) to allow for fibroblast cell division and, consequently, an increase in the number of fibroblasts bound to the matrix. Washing of the composition and either immediate use or shipping are
- 15 as described above for the sponge matrices.

- Compositions using Bio-Oss collagen, RESOLUT, and COLLACOTE as the matrix material have been used to heal extraction sockets of 18 patients after extraction of a tooth. In untreated extraction sockets, the subject's
- 20 fibroblasts migrate into the sockets approximately 10-14 days after extraction of the tooth and thus the healing process only begins at that time. However, by implanting the compositions containing the patient's fibroblasts into the sockets immediately after extraction of the
- 25 tooth (e.g., within 1-3 hours of the extraction), the healing process is initiated immediately. Furthermore, by implanting the compositions, shrinkage of the socket (due to collapse of the socket walls), which is generally approximately 30% in untreated sockets, is minimized. It
- 30 is particularly desirable that socket shrinkage be minimized in cases in which it is proposed to implant a dental prosthesis (e.g., a false tooth) at the site of the extraction at a later date. In all the compositions used for the above described procedures,
- 35 fibroblasts obtained from 1 mm punch biopsies of the

- 35 -

patients' gum tissues were used and the compositions were shipped to the dentists performing the procedures. The compositions were shaped by the dentist to fit into the extraction sockets. In general, the compositions after
5 shaping had an approximately cylindrical shape with a length of approximately 10 mm and a diameter of approximately 4 mm. At the time of implanting, fragments of the compositions were prepared for histological analysis. In all cases, fibroblast colonization of and
10 proliferation within the matrices was seen by light microscopy.

The above-mentioned advantages (i.e., facile healing and maintenance of socket volume) were seen in all cases. Furthermore, infection of the tissues
15 surrounding the socket, which is a frequently observed sequela of procedures in which extraction sockets are not treated, was not observed in any case. Compositions using Bio-Oss® collagen (2 patients) as the matrix had the advantage of persistence of the matrix material in
20 the sockets longer than those compositions in which the matrix material consisted of collagen only and thus provided physical support for the fibroblasts before full replacement of the matrix material with components endogenous to the patient.

25

Example III: An Injectable Composition for correcting
skin defects and augmenting tissue in a
Subject

As explained above, the present invention extends
30 to an injectable composition for correcting a defect in skin of a subject, or augmenting tissue of a subject, wherein the injectable composition comprises passaged autologous fibroblasts substantially free of immunogenic proteins, and a biodegradable, acellular injectable
35 filler material. Examples of skin defects that can be

- 36 -

treated with the present invention include scars, particularly facial scars resulting from trauma or acne, or wrinkles. Furthermore, the present invention can be used to augment tissue in a subject. For example, the
5 injectable composition of the present invention can be injected into the lips of a subject in order to make the lips larger and fuller. Fibroblasts are used in the present invention because they can be readily obtained and expanded, and are a cell type normally present
10 beneath the dermis. Fibroblasts taken from a biopsy of the gums, palate, or skin of the subject can be used in the present invention, and can readily obtained using procedures set forth above.

Initially, a dermal fibroblast culture is
15 initiated from a 1 to 5 mm full thickness biopsy specimen of the gums, palate or skin of the subject. Because of the phenomenon of allograft rejection, which is well known to transplantation surgeons and immunologists, it is essential that the cultured fibroblasts be
20 histocompatible with the host. Histocompatibility can be ensured by obtaining a biopsy of the subject to be treated and culturing the fibroblasts from this specimen.

Before the initiation of the culture, the biopsy is washed repeatedly with antibiotic and antifungal
25 agents. The specimen of dermis is then separated into small pieces. The pieces of the specimen are individually placed onto the dry surface of a tissue culture flask and allowed to attach for between about 5 and about 10 minutes, before a small amount of medium is
30 slowly added, taking care not to displace the attached tissue fragments. After about 48 hours of incubation, the flask is fed with additional medium. When a T-25 flask is used to start the culture, the initial amount of medium is about 1.5-2.0 ml. The establishment of a cell
35 line from the biopsy specimen ordinarily takes between

- 37 -

about 2 and 3 weeks, at which time the cells can be removed from the initial culture vessel for expansion.

During the early stages of the culture, it is desirable that the tissue fragments remain attached to the culture vessel bottom; fragments that detach should be reimplanted into new vessels. The fibroblasts can be stimulated to grow by a brief exposure to EDTA-trypsin, according to techniques well known to those skilled in the art. The exposure to trypsin is too brief to release the fibroblasts from their attachment to the culture vessel wall.

Immediately after the cultures have become established and are approaching confluence, samples of the fibroblasts can be processed for frozen storage, such as in liquid nitrogen. Presently, numerous methods for successfully freezing cells for later use are known in the art and are included in the present invention. The frozen storage of early rather than late passage fibroblasts is preferred because the number of passages in cell culture of normal human fibroblasts is limited.

The fibroblasts can be frozen in any freezing medium suitable for preserving fibroblasts. A medium consisting of about 70 % (v/v) growth medium, about 20 % (v/v) fetal bovine serum and about 10 % (v/v) dimethylsulfoxide (DMSO) can be used with good effect. DMSO can also be substituted with, for example, glycerol. Thawed cells can be used to initiate secondary cultures to obtain suspensions for use in the same subject without the inconvenience of obtaining a second specimen.

Any tissue culture technique that is suitable for the propagation of dermal fibroblasts from biopsy specimens may be used to expand the cells to practice the invention. Techniques well known to those skilled in the art can be found in R.I. Freshney, Ed., ANIMAL CELL CULTURE: A PRACTICAL APPROACH (IRL Press, Oxford,

- 38 -

England, 1986) and R.I. Freshney, Ed., CULTURE OF ANIMAL CELLS: A MANUAL OF BASIC TECHNIQUES, Alan R. Liss & Co., New York, 1987), which are hereby incorporated by reference.

5 The medium can be any medium suited for the growth of primary fibroblast cultures. The medium can be supplemented with human or non-human serum in an amount of between about 0.0 % and about 20 % (v/v) to promote growth of the fibroblasts. Higher concentrations of
10 serum promote faster growth of the fibroblasts. An example of medium having application herein comprises glucose DMEM supplemented with about 2 mM glutamine, about 110 mg/L sodium pyruvate, about 10 % (v/v) fetal bovine serum and antibiotics ("complete medium"), wherein
15 the concentration of glucose ranges from approximately 1,000 mg/L of medium to 4,500 mg/L of medium, and preferably is 4,500 mg/L. Fibroblasts can also be expanded in a serum-free medium.

 Autologous fibroblasts can be passaged into new
20 flasks by trypsinization. For expansion, individual flasks are split 1:3. Triple bottom, T-150 flasks, having a total culture area of 450 cm² are suitable for the practice of the invention. A triple bottom T-150 flask can be seeded with about 1X10⁶ to about 3X10⁶ cells
25 and has a capacity to yield about 8X10⁶ to about 1.3X10⁷ cells. When the capacity of the flask is reached, which typically requires about 5-7 days of culture, the growth medium is replaced by serum-free medium; thereafter the cells are incubated, i.e., held at between about 30° C
30 and about 37.5° C, for at least 2 hours. The incubation of the cells in serum-free medium substantially removes from the cells proteins that are derived from the fetal bovine serum which, if present, can elicit an immune response in the subject. In a preferred embodiment,
35 serum-free medium comprises glucose DMEM supplemented

- 39 -

with about 2 mM glutamine, and about 110 mg/L sodium pyruvate, wherein the concentration of glucose is approximately 1,000 mg/L of medium to approximately 4,500 mg/L of medium, and preferably 4,500 mg/L of medium.

5 At the end of the incubation in serum-free medium, the cells are removed from the tissue culture flask by trypsin-EDTA; washed extensively by centrifugation and resuspension; and suspended for subsequent use in an injectable composition of the present invention, or for
10 injection into the subject. Five triple bottom T-150 flasks, grown to capacity, yield about 8×10^6 to about 1.3×10^7 cells which are sufficient to make up about 1.2 to 1.4 ml of cell suspension.

 The cells can be transported at 4° C so long as
15 they are injected within 24-48 hours of their suspension. The cells can be suspended in an appropriate physiological solution with appropriate osmolarity and tested for pyrogens and endotoxin levels, except for the absence of phenol red pH indicator, and the replacement
20 of the fetal bovine serum by the subject's serum for such transportation (transport medium). In another embodiment the cells can be suspended in Krebs-Ringer solution comprising 5 % dextrose or any other physiological solution. The cells can be aspirated and injected into
25 the transport medium.

 The volume of saline or transport medium in which the cells are suspended is related to such factors as the number of fibroblasts the practitioner desires to inject, the extent of the defects to the subject's skin that are
30 to be corrected, the size or number of the defects that are to be corrected, and the urgency of the subject's desire to obtain the results of the treatment. Moreover, the practitioner can suspend the cells in a larger volume of medium and inject correspondingly fewer cells at each
35 injection site. In an injectable composition of the

- 40 -

present invention, the passaged autologous fibroblasts of the present invention are mixed with a biodegradable, acellular injectable filler material in a ratio of approximately 1:1 by volume.

5 Biodegradable, acellular injectable filler material

As explained above, an injectable composition of the present invention also comprises a biodegradable, acellular injectable filler material. Numerous types of biodegradable, acellular injectable filler materials are presently available and have applications in the present invention. More specifically, the filler material can be comprised endogenous proteins, such as any type of collagen from the subject. An example of such a filler is "AUTOLOGEN" produced by Collagenesis, Inc.

15 "AUTOLOGEN" is a dispersion of autologous dermal collagen fibers from the subject, and should not elicit an immune response. In order to obtain "AUTOLOGEN" for the subject, a specimen of tissue is obtained from the subject and forwarded to Collagenesis, Inc., where it is turned into "AUTOLOGEN". Approximately a one and a half square inch of tissue yields one cubic centimeter (cc) of "AUTOLOGEN". After "AUTOLOGEN" has been prepared, its concentration can be adjusted depending upon the amount needed to correct defects in the subject's skin, or

20 augment tissue in the subject. In particular, the concentration of "AUTOLOGEN" in the dispersion can be at least about 25 mg/L.

Another example of filler material comprises exogenous proteins, such as any type of collagen.

30 Presently, numerous collagen products are commercially available and have applications in the present invention. Examples of such products are reconstituted bovine collagen products commercially available including, but not limited to, "ZYDERM I", "ZYDERM II" and "ZYPLAST",

- 41 -

which comprise reconstituted bovine collagen fibers cross-linked with glutaraldehyde. These three products, which have been approved by the U.S. Food and Drug Administration (FDA) for treating wrinkle lines and depressed scars since 1981, are produced by the Collagen Corporation of Palo Alto, CA.

Other examples of filler materials having applications in the present invention include, but are not limited to, solubilized gelatin, polyglycolic acid, or cat gut sutures. One particular is a gelatin matrix implant sold under the mark "FIBRIL", which comprises porcine gelatin powder plus o-aminocaproic acid, which are dispersed in a 0.9 % (by volume) sodium chloride solution and an aliquot of plasma from the subject, mixed in a 1:1 ratio by volume.

Methods for correcting a defect in skin
or augmenting tissue

Furthermore, the present invention extends to methods for correcting a defect in skin of a subject, such as scars or wrinkles, or augmenting tissue in the subject, particularly facial tissue, e.g., augmenting the tissue of lips to make the lips appear fuller. One such method of the present invention comprises injecting an effective amount of an injectable composition comprising autologous fibroblasts substantially free of immunogenic proteins and a biodegradable, acellular injectable filler material, into the subject at the site of the skin defect or desired tissue augmentation so that regeneration of tissue at the site is promoted.

Another method encompassed by the present invention comprises the steps of injecting passaged autologous fibroblasts substantially free of immunogenic proteins into the subject at a site of a skin defect or desired tissue augmentation, and injecting a biodegradable, acellular injectable filler material into

- 42 -

the site. Preferably, the duration between injecting the autologous fibroblasts into the subject and injecting the biodegradable acellular injectable filler into the subject is about two weeks.

5 Passaged, autologous fibroblasts substantially free of immunogenic proteins can be readily obtained using procedures set forth above. Furthermore, the various types of biodegradable, acellular filler materials, which have been described in detail above,
10 have applications in the methods described herein.

 Injections set forth in the present invention are typically carried out with a hypodermic syringe having a syringe chamber, a piston disposed therein, an orifice communicating with the chamber, and a hypodermic needle
15 is fixed to the orifice. The size of the needle used in a method for correcting a defect in skin of a subject or augmenting tissue in the subject ranges from approximately 30 gauge to approximately 27 gauge.

 Initially, the tissue to be injected is prepped
20 with alcohol and stretched to give a taut surface. After the tissue to be injected has been prepped, a syringe is filled with an injectable composition of the present invention, if the first method described above is to be used, or with passaged, autologous fibroblasts
25 substantially free of immunogenic proteins if the second method for correcting skin defects or augmenting tissue is used. The needle is inserted into the tissue as superficially as possible, and the orientation of the bevel is not critical to the success of this method of
30 present invention. The actual injection is made by gentle pressure until a slight blanch is seen in the injected tissue. Multiple serial injections can be made. Approximately two weeks after passaged autologous fibroblasts are injected into the subject's skin, an
35 equal volume of biodegradable, acellular filler is

- 43 -

injected using the same procedure as described above into the same location where the passaged autologous fibroblasts were previously injected.

The present invention is not to be limited in
5 scope by the specific embodiments described above, which are intended as illustrations of aspects of the invention. Here functionally equivalent methods and components are within the scope of the invention. Indeed, various modifications of the invention, in
10 addition to those shown and described herein, will become apparent to those skilled in the art from the foregoing description. Such modifications are intended to fall within the scope of the appended claims. Moreover, all cited references are, hereby, incorporated by reference.

- 44 -

WHAT IS CLAIMED IS:

1. A composition for promoting regeneration of tissue that has degenerated in a subject as a result of a disease or disorder in said subject, wherein said
5 composition comprises:
a biodegradable acellular matrix; and
autologous fibroblasts substantially free of immunogenic proteins, wherein said autologous fibroblasts are integrated within said acellular matrix.
- 10 2. The composition of Claim 1, wherein said biodegradable acellular matrix comprises exogenous proteins.
3. The composition of Claim 2, wherein said biodegradable acellular matrix comprises any type of
15 collagen.
4. The composition of Claim 2, wherein said biodegradable acellular matrix comprises any type of collagen and glycosaminoglycans, cross-linked with glutaraldehyde.
- 20 5. The composition of Claim 1, wherein said biodegradable acellular matrix comprises one or more substances selected from the group consisting of gelatin, polyglycolic acid, cat gut, demineralized bone, and hydroxyapatite.
- 25 6. The composition of Claim 5, wherein said biodegradable acellular matrix comprises gelatin, polyglycolic acid, or cat gut.
7. The composition of Claim 1, wherein said disease or disorder in said subject comprises defects of an oral
30 mucosa, trauma to an oral mucosa, periodontal disease,

- 45 -

diabetes, cutaneous ulcers, venous stasis, scars of skin, or wrinkles of skin.

8. The composition of Claim 7, wherein said disease or disorder is periodontal disease, and said periodontal
5 disease comprises periodontal degeneration, gingivitis, or non-healing wounds of a palatal mucosa or a gingival mucosa.

9. The composition of Claim 1, wherein said autologous fibroblasts are from gums, palate, or skin of
10 said subject.

10. A method for making a composition for regenerating tissue that has degenerated in a subject as a result of a disease or disorder in said subject, wherein said method comprises:

15 providing a suspension of autologous, passaged fibroblasts substantially free of immunogenic proteins;
providing a biodegradable acellular matrix;
incubating said suspension of autologous passaged fibroblasts with said biodegradable acellular matrix such
20 that said autologous passaged fibroblasts integrate within said biodegradable acellular matrix, to form a composition for promoting the regeneration of tissue.

11. The method of Claim 10, wherein said biodegradable acellular matrix comprises exogenous proteins.

25 12. The method of Claim 11, wherein said acellular matrix comprises any type of collagen.

13. The method of Claim 12, wherein said acellular matrix comprises any type of collagen cross-linked glycosaminoglycans, cross-linked with glutaraldehyde.

- 46 -

14. The method of Claim 9, wherein said biodegradable
acellular matrix comprises one or more substances
selected from the group consisting of gelatin,
polyglycolic acid, cat gut, demineralized bone, and
5 hydroxyapatite.

15. The method of Claim 14, wherein said biodegradable
acellular matrix comprises gelatin, polyglycolic acid, or
cat gut.

16. The method of Claim 10, wherein said disease or
10 disorder in said subject comprises defects of an oral
mucosa, trauma to an oral mucosa, periodontal disease,
diabetes, cutaneous ulcers, venous stasis, scars of skin,
or wrinkles of skin.

17. The method of Claim 16, wherein said disease or
15 disorder is periodontal disease, and said periodontal
disease comprises periodontal degeneration, gingivitis,
or a non-healing wound of a palatal mucosa or a gingival
mucosa.

18. The method of Claim 10, wherein the step of
20 providing a suspension of autologous, passaged
fibroblasts substantially free of immunogenic proteins
comprises:

collecting a biopsy of dermis of said subject;
separating dermal autologous fibroblasts from said
25 biopsy;

culturing said dermal autologous fibroblasts in a
culture medium comprising between 0.0% and about 20%
human or non-human serum;

incubating said dermal autologous fibroblasts in a
30 serum-free medium for at least about 2 hours between
about 30° C and 37.5° C; and

- 47 -

exposing said incubated dermal autologous fibroblasts to a proteolytic enzyme so as to suspend fibroblasts.

19. The method of Claim 18, wherein the step of
5 collecting a biopsy of dermis further comprises collecting a biopsy from gums, palate or skin of said subject.

20. A method for promoting regeneration of tissue that has been damaged in a subject as a result of a disease or
10 disorder in said subject, wherein said method comprises:

- a) providing a composition comprising of autologous, passaged fibroblasts substantially free of immunogenic proteins integrated into a biodegradable acellular matrix;
- 15 b) identifying a site of tissue degeneration or a defect;
- c) placing the composition on the area of tissue degeneration or defect so that said tissue is augmented.

21. A method for promoting regeneration of tissue that
20 has been damaged in a subject, wherein the method comprises:

- a) providing a pharmaceutical composition comprising autologous passaged fibroblasts substantially free of immunogenic proteins, and a pharmaceutically
25 acceptable carrier thereof;
- b) identifying a site of tissue degeneration or defect;
- c) injecting a therapeutically effective amount of the pharmaceutical composition subadjacent to the
30 tissue degeneration or defect until a slight blanch is seen in the injected tissue.

- 48 -

22. The method of Claim 21, wherein said disease or disorder in said subject comprises defects of an oral mucosa, trauma to an oral mucosa, periodontal disease, diabetes, cutaneous ulcers, venous stasis, scars of skin, or wrinkles of skin.

23. The method of Claim 22, wherein said disease or disorder is periodontal disease, wherein said periodontal disease comprises periodontal degeneration, gingivitis, or a non-healing wound of a palatal mucosa or a gingival mucosa.

24. The method of Claim 21, wherein the step of providing a pharmaceutical composition comprises:

- a) taking a biopsy of dermis comprising fibroblasts from a subject;
- 15 b) separating fibroblasts from said biopsy so as to provide fibroblasts substantially free of extracellular matrix and non-fibroblast cells;
- c) placing the fibroblasts in a culture medium comprising between 0.0% and about 20% serum in order to grow fibroblasts;
- 20 d) incubating the fibroblasts in a serum-free medium for at least 2 hours at between about 30° C and about 37.5° C to form passaged fibroblasts;
- e) exposing the passaged fibroblasts to a proteolytic enzyme so as to suspend the passaged fibroblasts; and
- 25 f) adding a pharmaceutically acceptable carrier to the suspended passaged fibroblasts to form the pharmaceutical composition.

30 25. The method of Claim 24, wherein biopsy is taken from gums, palate, or skin of the subject.

- 49 -

26. The method of Claim 24 wherein said proteolytic enzyme is trypsin.

27. A pharmaceutical composition for promoting regeneration of tissue that has been damaged in a subject
5 comprising passaged autologous fibroblasts substantially free of immunogenic proteins, and a pharmaceutically acceptable carrier thereof.

28. The pharmaceutical composition of Claim 27, wherein said passaged autologous fibroblasts comprise
10 fibroblasts of the gums, palate, or skin.

29. An injectable composition for correcting a defect in skin of a subject, or augmenting tissue of a subject, said injectable composition comprising:

- a) autologous fibroblasts substantially free of
15 immunogenic proteins; and
- b) a biodegradable, acellular injectable filler material.

30. The injectable composition of Claim 29, wherein said autologous fibroblasts are from gums, palate of skin
20 of said subject.

31. The injectable composition of Claim 29, wherein said biodegradable, acellular injectable filler material comprises endogenous proteins.

32. The injectable composition of Claim 31, wherein
25 said biodegradable acellular injectable filler material comprises an injectable dispersion of autologous collagen fibers.

- 50 -

33. The injectable composition of Claim 32, wherein the concentration of autologous collagen fibers in said injectable dispersion is at least 24 mg/ml.

34. The injectable composition of Claim 29, wherein
5 said biodegradable acellular injectable filler material comprises exogenous proteins.

35. The injectable composition of Claim 34, wherein said exogenous proteins comprise any type of collagen.

36. The injectable composition of Claim 35, wherein
10 said biodegradable, acellular injectable filler material comprises reconstituted bovine collagen fibers cross-linked with glutaraldehyde.

37. The injectable composition of Claim 29, wherein said biodegradable, acellular injectable filler material
15 comprises solubilized gelatin, polyglycolic acid, or cat gut.

38. The injectable composition of Claim 37, wherein said biodegradable, acellular injectable filler material comprises porcine gelatin powder and amino caproic acid
20 dispersed in sodium chloride solution and an aliquot of plasma from said subject.

39. The injectable composition of Claim 38, wherein the ratio of said sodium chloride solution and said aliquot of serum is 1:1 by volume.

25 40. The injectable composition of Claim 39, wherein said sodium chloride solution comprises 0.9% sodium chloride by volume.

- 51 -

41. A method for correcting a defect in skin of a subject, or augmenting tissue of a subject, the method comprising injecting an effective amount of an injectable composition comprising autologous fibroblasts
- 5 substantially free of immunogenic proteins and a biodegradable, acellular injectable filler material, into the subject at the site of the skin defect or desired tissue augmentation so that regeneration of tissue at the site is promoted.
- 10 42. A method for correcting a defect in skin of a subject, or augmenting tissue of a subject, the method comprising the steps of:
- a) injecting autologous fibroblasts substantially free of immunogenic proteins into the
- 15 subject at a site of a skin defect or desired skin augmentation; and
- b) injecting a biodegradable, acellular injectable filler material into the site.
43. The method for correcting a defect in skin of a
- 20 subject, or augmenting tissue of a subject of Claim 42, wherein the duration between injecting the autologous fibroblasts into the subject and injecting the biodegradable acellular injectable filler into the subject is about two weeks.
- 25 44. The method of Claim 42, wherein the autologous fibroblasts are from gums, palate of skin of the subject.
45. The method of Claim 42, wherein the biodegradable, acellular injectable filler material comprises endogenous proteins.

- 52 -

46. The method of Claim 45, wherein the biodegradable, acellular injectable filler material comprises an injectable dispersion of autologous collagen fibers.

47. The method of Claim 46, wherein the concentration
5 of autologous collagen fibers in the injectable dispersion is at least 24 mg/ml.

48. The method of Claim 42, wherein the biodegradable, acellular injectable filler material comprises exogenous proteins.

10 49. The method of Claim 48, wherein the exogenous proteins comprise any type of collagen.

50. The method of Claim 49, wherein the biodegradable, acellular injectable filler material comprises reconstituted bovine collagen fibers cross-linked with
15 glutaraldehyde.

51. The method of Claim 42, wherein the biodegradable, acellular injectable filler material comprises solubilized gelatin, polyglycolic acid, or cat gut.

52. The method of Claim 51, wherein the biodegradable,
20 acellular injectable filler material comprises porcine gelatin powder and aminocaproic acid dispersed in sodium chloride solution, and an aliquot of plasma from the subject.

53. The method of Claim 52, wherein the ratio of
25 sodium chloride solution to the aliquot of serum is 1:1 by volume.

- 53 -

54. The method of Claim 53, wherein the sodium chloride solution comprises 0.9% sodium chloride by volume.

55. The method of Claim 42, wherein the ratio of autologous fibroblasts substantially free of immunogenic proteins to biodegradable, acellular injectable filler material is approximately 1:1 by volume.

56. The composition of Claim 1, wherein said biodegradable acellular matrix comprises anorganic bone.

10 57. The composition of Claim 56, further comprising collagen.

58. The method of Claim 10, wherein said biodegradable acellular matrix comprises anorganic bone.

59. The method of Claim 58, said biodegradable
15 acellular matrix further comprising collagen.

60. The method of Claim 20, wherein said biodegradable acellular matrix comprises exogenous proteins.

61. The method of Claim 60, wherein said biodegradable
20 acellular matrix comprises any type of collagen.

62. The method of Claim 60, wherein said biodegradable acellular matrix comprises any type of collagen and glycosaminoglycans, cross-linked with glutaraldehyde.

63. The method of Claim 20, wherein said biodegradable
25 acellular matrix comprises one or more substances selected from the group consisting of gelatin,

- 54 -

polyglycolic acid, cat gut sutures, demineralized bone, and hydroxyapatite.

64. The method of Claim 20, wherein said disease or disorder in said subject comprises defects of an oral
5 mucosa, trauma to an oral mucosa, periodontal disease, diabetes, cutaneous ulcers, venous stasis, scars of skin, or wrinkles of skin.

65. The method of Claim 64, wherein said disease or disorder is periodontal disease, and said periodontal
10 disease comprises periodontal degeneration, gingivitis, or non-healing wounds of a palatal mucosa or a gingival mucosa.

66. The method of Claim 20, wherein said autologous fibroblasts are from gums, palate, or skin of said
15 subject.

67. The method of Claim 20, wherein said biodegradable acellular matrix comprises anorganic bone.

68. The method of Claim 67, said biodegradable acellular matrix further comprising collagen.

20 69. The method of Claim 64, wherein said trauma to the oral mucosa is an extraction socket resulting from an extraction of a tooth.

70. The composition of Claim 7, wherein said trauma to the oral mucosa is an extraction socket resulting from an
25 extraction of a tooth.

- 55 -

71. The method of Claim 16, wherein said trauma to the oral mucosa is an extraction socket resulting from an extraction of a tooth.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US99/11283

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : A61F 2/00, 2/02, 2/10, 13/00; A61K 38/00, 38/17; A61M 31/00

US CL : 424/422, 423, 426, 428; 514/21; 530/356; 604/51; 623/11; 623/15

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/422, 423, 426, 428; 514/21; 530/356; 604/51; 623/11; 623/15

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 5,166,187 A (COLLOMBEL et al.) 24 November 1992, entire document.	1-71
Y	US 5,591,444 A (BOSS, JR.) 07 January 1997, entire document.	1-71
Y	US 5,660,850 A (BOSS, JR.) 26 August 1997, entire document.	1-71
Y	US 5,665,372 A (BOSS, JR.) 09 September 1997, entire document.	1-71
Y	US 5,711,957 A (PATAT et al.) 27 January 1998, entire document.	1-71



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
B earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Z* document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

19 JULY 1999

Date of mailing of the international search report

10 SEP 1999

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/11283

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y,P	LAMME et al. Living skin substitutes: Survival and function of fibroblasts seeded in a dermal substitute in experimental wounds. J. Investigative Dermatology. December 1998, Vol. 111, No. 6, pages 989-995, entire document.	1-71

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/11283

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS

STN (file: medicine)

search terms: bone, collagen, silicone, fat, gelatin, polymethylacrylate, glycosaminoglycan, cat(w)gut, polyglycolic acid, hydroxyapatite, biodegradable, matrix, acellular, biocompatible, regenerat?, fibroblast#, autologous